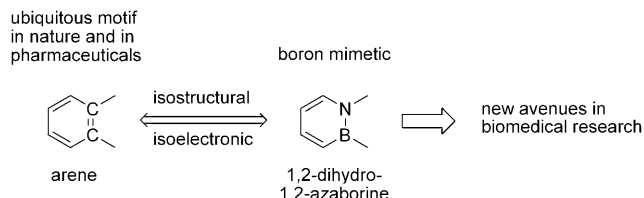


Boron Mimetics: 1,2-Dihydro-1,2-azaborines Bind inside a Nonpolar Cavity of T4 Lysozyme**

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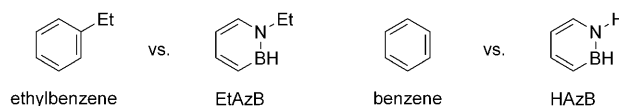
The element boron has not received much attention in biomedical applications compared to its neighbors in the periodic table carbon, nitrogen, and oxygen. Arguably, this situation might be due to the apparent insignificance of boron in nature's evolution of life.^[1,2] Boron has, however, useful elemental and chemical features that include nuclear spin, large cross section for neutron capture, and Lewis acidity. If boron could be incorporated into biologically relevant molecules,^[3] it might benefit biomedical research by being used as a marker,^[4] as a new pharmacophore,^[5] or in cancer therapy.^[6] We are interested in synthetic approaches that will allow the incorporation of boron into such molecules^[7] with minimal perturbation of their structures. 1,2-Dihydro-1,2-azaborines (abbreviated as 1,2-azaborines) serve as a unique structural platform to accomplish this goal because of their isostructural relationship with arenes, a ubiquitous motif in living organisms and in pharmaceuticals. Furthermore, the bonding of arenes with cations and other arenes through cation- π ^[8] and π - π ^[9] interactions have been demonstrated to be vital in biological systems. Thus, the broad utility and fundamental importance of arenes in biomedical research combined with the unique elemental and chemical features of boron and the potential of expanding the diversity of arene structures through CC/BN isosterism^[10] make 1,2-azaborines attractive targets for biomedical investigation (Scheme 1).

Dewar et al. and White pioneered the chemistry of monocyclic and ring-fused polycyclic 1,2-azaborine derivatives in the 1960s.^[11] Since 2000, contributions by the groups of Ashe,^[12] Piers,^[13] and Paetzold^[14] have further advanced the preparation of novel BN heterocycles and sparked a renewed interest in the chemistry and properties of these compounds.^[15] We have recently developed synthetic methods that expand the scope of accessible 1,2-azaborines^[16] and enable the preparation of the long-sought parent compound



Scheme 1. 1,2-Azaborines as boron-containing arene mimetics.

of this family of heterocycles.^[17] We have also shown through X-ray crystallographic studies that 1,2-azaborines possess delocalized structures consistent with aromaticity.^[18] While the development of a versatile synthetic toolbox for 1,2-azaborines has been improved, the investigation of these compounds in a biological context has remained elusive. Herein, we provide the first experimental evidence for the interaction of 1,2-azaborines with a biological system. Specifically, we describe the binding of *N*-ethyl-1,2-azaborine and the parent 1,2-azaborine (hereafter EtAzB and HAzB, respectively) inside an engineered nonpolar cavity of T4 lysozyme, which was investigated using single-crystal X-ray crystallography.



The mutation Leu99→Ala (L99A) in T4 lysozyme^[19] creates a disc-shaped apolar cavity with a volume of approximately 160 Å³. Although this cavity is internal, it readily binds various arenes and other ligands.^[20,21] We are interested in addressing the following questions: 1) Do EtAzB and HAzB bind to L99A lysozyme? 2) If so, do they bind in the same fashion as their all-carbon analogues?

EtAzB^[16] and HAzB^[17] were prepared according to our reported procedure. L99A lysozyme was purified and crystallized as described.^[19,22] The protein crystals and their soaking solution (2.2 M sodium/potassium phosphate, pH 6.9, 50 mM 2-mercaptoethanol, 50 mM hydroxyethylhydrosulfide) were then deoxygenated. Complexes of L99A with EtAzB, ethylbenzene, HAzB, and benzene were prepared by vapor diffusion of the respective ligand into the protein crystals in sealed oxygen-free glass tubes at 277 K for three days. Diffraction data (see the Supporting Information) were collected from frozen crystals (100 K) to 1.25 Å resolution.^[24] Crystallographic refinement was carried out using the CCP4^[23] package.

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Figure 1 shows the $F_o - F_c$ electron density maps for EtAzB and ethylbenzene bound in the L99A cavity. As is immediately apparent, both ligands bind in a similar fashion. In both cases the aromatic ring occupies the classic benzene

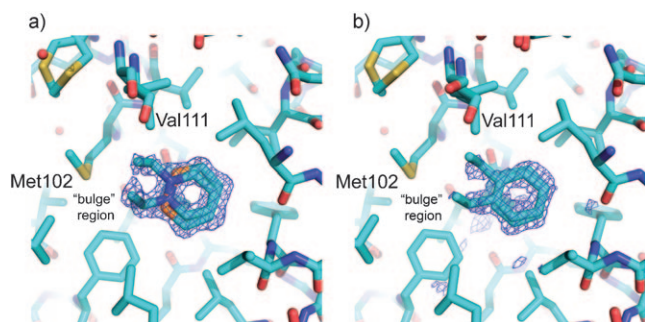


Figure 1. Binding of a) EtAzB and b) ethylbenzene in the T4 lysozyme L99A cavity. The difference maps (blue) have coefficients $(F_o - F_c)\exp(i\alpha_c)$, where F_o is the amplitude observed for crystals of the complex and F_c and α_c are the amplitudes and phases, respectively, calculated from the refined model prior to including the ligand. The maps are contoured at 3σ and 2σ for EtAzB and ethylbenzene, respectively, at a resolution of $1.25\text{ \AA}$. The nitrogen atom on the 1,2-azaborine ring of EtAzB is colored blue. The sites for the boron atoms could not be specified unambiguously (see text).

binding site.^[19,20] Also for both ligands, the ethyl group occupies two alternative positions. In one of these positions the ethyl moiety protrudes into the “bulge” region of the cavity and makes a van der Waals contact (ca. $3.8\text{ \AA}$) with the sulfur atom of Met102. In the alternative conformation, the ethyl group still maintains a van der Waals contact (ca. $3.8\text{ \AA}$) with Met102 but occupies space between Met102 and Val111. This situation causes the side chain of Val111 to rotate approximately 120° into a different conformer. The crystallographic refinement suggests that EtAzB binds to L99A with essentially 100% occupancy, whereas the occupancy for ethylbenzene is about 60%. In a prior lower-resolution study of ethylbenzene binding to L99A at room temperature,^[20] the geometry of binding was similar to that shown in Figure 1 b, except that the “bulge” region binding mode was predominant.

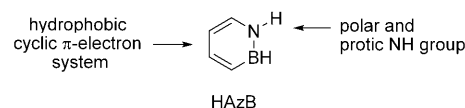
The difference electron density (Figure 1a) and the subsequent crystallographic refinement indicate that the atoms in the azaborine ring as well as the benzylic carbon atom of the ethyl group are coplanar. This result is in agreement with prior observations using small-molecule crystallography.^[16] The present analysis suggests that EtAzB is stable in the cavity, at least during the preparation time (three days at room temperature) plus the much shorter X-ray data collection time at 100 K . There is no obvious electron density suggesting that the ligand might have chemically modified some site on the protein.

Although EtAzB and ethylbenzene bind similarly, there are some differences. As can be seen in Figure 1 b, the phenyl rings of ethylbenzene in its two alternative modes of binding superimpose within about $0.2\text{ \AA}$. For EtAzB, however (Figure 1 a), the azaborine rings are offset relative to each other

by about $0.6\text{ \AA}$. The difference between EtAzB and ethylbenzene may be due in part to the C–B bond (ca. $1.5\text{ \AA}$), which is approximately $0.1\text{ \AA}$ longer than the aromatic C–C bond.^[18] It should be noted that owing to the alternative positions occupied by EtAzB it is not possible to reliably identify the specific site(s) occupied by the boron atom. No putative close contacts between the N or B atoms of EtAzB and the surrounding atoms of the protein are apparent. This finding suggests that polar interactions do not have a dominant effect on the binding of this ligand, which again is consistent with the previous observation that the 1,2-azaborine ring is aromatic and has considerable hydrophobicity.

In contrast to EtAzB, the parent 1,2-azaborine HAzB contains a polar and protic N–H bond that readily undergoes isotope exchange in the presence of $[D_4]$ methanol.^[17] Consistent with this protic bond, HAzB has a significantly lower R_f value of 0.4 on silica gel (using pentane as eluent) than benzene under the same conditions ($R_f \approx 1.0$). It has been shown that a number of polar ligands, including pyridine, phenol, and aniline do not bind significantly to the L99A cavity.^[20a] Thus, efficient binding of HAzB to the hydrophobic cavity of L99A was questionable owing to the dichotomous characteristics exhibited by HAzB (Scheme 2).

Crystals of L99A lysozyme in the presence of HAzB were prepared and analyzed in the same manner as for EtAzB.



Scheme 2. The dichotomy of the parent HAzB.

Gratifyingly, we observed that the resultant difference electron density map (Figure 2a) has a strong disc-shaped feature clearly indicating the binding of HAzB in the cavity. As a control, the binding of benzene, previously shown at room temperature,^[19b] was redetermined under the low-temperature conditions used for HAzB and EtAzB (Figure 2 b). There is essentially no change in the cavity (less than $0.1\text{ \AA}$) when either HAzB or benzene is bound.

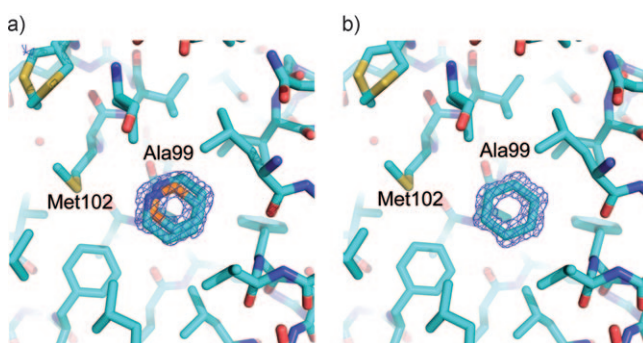


Figure 2. Binding of HAzB (a) and benzene (b) in the T4 lysozyme L99A cavity. The difference electron density maps (blue), both contoured at the 3σ level, are calculated from coefficients $(F_o - F_c)\exp(i\alpha_c)$. The nitrogen atom on the 1,2-azaborine ring is colored blue. Boron atom sites are included (orange).

The electron density for the benzene complex (Figure 2b) suggests that the six carbon atoms occupy well-defined sites. For HAZB, however, the density in the vicinity of Met102 and Val111 suggests that HAZB may bind in two conformations, the first with the nitrogen atom 4.0 Å from the sulfur atom of Met102 and the second “flipped over” with the nitrogen atom 3.7 Å from the carbonyl oxygen of Ala99. In neither case is there any indication of a strong hydrogen-bonding interaction. This finding is consistent with HAZB having apolar rather than polar characteristics inside this specific protein environment. Moreover, HAZB appears to bind to L99A lysozyme with essentially 100% occupancy. This structure of HAZB bound to L99A represents the first crystallographic characterization of the parent 1,2-azaborine in its metal-free form.

In summary, we furnished the first experimental evidence for the binding of 1,2-azaborines, including that of the parent HAZB, to a biological system. We demonstrate that unnatural 1,2-azaborines bind inside the cavity of T4 lysozyme L99A, and that they bind in a very similar fashion to their all-carbon isosteres. The observed binding of the parent HAZB is consistent with its nonpolar characteristics despite its protic NH functionality. Our studies provide a proof of concept that 1,2-azaborines can serve as boron-containing, hydrophobic arene mimics in a biological context. In view of the abundance and importance of arene structures in biomedical applications and the unique elemental and chemical features of boron, we believe that our studies will encourage the development of 1,2-azaborines in biologically relevant applications.

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